DNA after Trizol Prep.

First, follow the steps to isolate RNA. At the end of the procedure, you are left with the trizol + interphase (supernatant was taken for RNA).

1. Mix Trizol with Interphase by quick vortex, plus small spin to get “stuff” off of the cap.
2. Pipette trizol/interphase mix into a new tube.
3. Add .3ml of 100% ETOH.
4. Vortex briefly. Sit at room temp for 2-3 min.
5. Spin at 2000 x g for 5 min. NOTE: this is a much slower spin than used for RNA. Check the centrifuge for proper setting.
6. Wash Pellet with Trisodium citrate/ETOH solution
   1. .1M Trisodium Citrat in 10% ETOH.
7. Incubate pellet at room temp for 30min.
8. Repeat spin step #5
9. Dissolve pellet in 600 ul of 2X cTAB.
   1. Incubate at 65-75oC might help with re-dissolving the pellet.
   2. I’ve also tried by re-dissolving in 600ul of H2O, but you than you need to do an NaAC/ETOH precipitation
10. Add 600ul of 25:24:1 or Phenole:Chloroform:Isoamyal alcohol solution
11. Mix by vortex, and sit at room temp for 5 min.
12. Spin at 12000 x g for 10 minutes
13. Take off supernatant and add equal volume of 100% ETOH
14. Vortex and sit at room temp for 5 min.
15. Spin 15min to pellet DNA
16. Wash DNA pellet with 75% ETOH
17. Spin for 5 min to pellet DNA
18. Air dry to 10-30min.
19. Re-dissolve in water (or TE) and use as needed.